

Spectroscopic Studies of Biologically Active Organotin(IV) Derivatives of 2-[N-(2,4,6-Tribromophenylamido)propanoic Acid

K. Shahid^a, S. Shahzadi^b and S. Ali^{a,*}

^aDepartment of Chemistry, Quaid-i-Azam University, Islamabad-45320, Pakistan

^bDepartment of Chemistry, GC University, Faisalabad, Pakistan

(Received 12 October 2007, Accepted 17 December 2007)

Herein we describe the synthesis and characterization of compounds having the formulae R_2SnL_2 and R_3SnL , where R = Me, *n*-Bu, Ph and *n*-Oct and L = 2-[N-(2,4,6-tribromophenylamido)propanoic acid. All the complexes have been characterized by various spectroscopic methods (IR and 1H , ^{13}C , ^{119}Sn NMR), elemental analysis, mass spectrometry and physical data. These compounds were also screened for their biological activity and found some encouraging results.

Keywords: Organotin(IV) carboxylates, NMR, IR, Mass, Biological activity

INTRODUCTION

Tin played a full part in the great increase of activity in organometallic chemistry, which started in about 1949 and this was stimulated by the discovery of variety of applications [1]. The considerable development over recent decades in the use of organotin compounds as reagents or intermediate in inorganic synthesis has promoted the preparation of many new organotin compounds and the developments of new, rapid and convenient synthetic procedures [2,3].

The most significant recent developments in organotin carboxylates is because of biological activity associated with these compounds; thus, there has been a large increase in reports on their synthesis, structure elucidation and biological activity-structure relationships [4-6].

Recently, considerable attention has been paid to triorganotin(IV) derivatives having high *in vitro* antifungal activities against some medically important fungi. The low aqueous solubility of organotin compounds is a limiting factor

in further research of their use in medicine [7]. On the basis of the known electron-acceptor properties of these compounds, it can be proposed that their toxicity is related to their interaction with electron donor groups in biologically important molecules. Referring to various applications of organotin carboxylates and continuation of our studies of biologically active organotin(IV) derivatives of substituted anilines [8,9], here we synthesized some organotin(IV) derivatives of 2-[N-(2,4,6-tribromophenylamido)propanoic acid (Fig. 1). These complexes were characterized by elemental analysis, infrared, multinuclear NMR (1H , ^{13}C , ^{119}Sn) and mass spectrometry. Their biological activity data has also been reported.

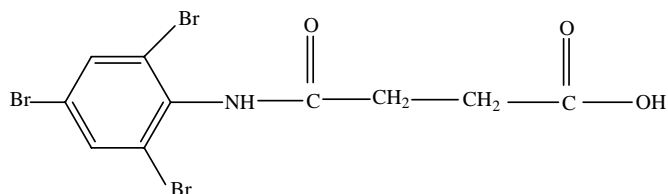


Fig. 1. Chemical structure of 2-[N-(2,4,6-tribromophenylamido)propanoic acid.

*Corresponding author. E-mail: drsa54@yahoo.com

EXPERIMENTAL

Material and Methods

All the chemicals were of analytical grade and used without further purification. Melting points were determined in capillary tubes using a MP-D Mitamura Riken Kogyo (Japan) electrothermal melting point apparatus and are uncorrected. Infrared absorption spectra were recorded as KBr (4000-400 cm^{-1}) pellets on Bio-Rad FT-IR spectrometer.

The ^1H , ^{13}C and ^{119}Sn NMR spectra were recorded on a Bruker AM 250 spectrometer (Germany), using CDCl_3 as an internal reference [δ of $^1\text{H}(\text{CDCl}_3) = 7.25$ and δ of $^{13}\text{C}(\text{CDCl}_3) = 77.0$]. ^{119}Sn NMR spectra were obtained using Me_4Sn as external reference [δ of $^{119}\text{Sn} = 37.290665$]. Mass spectral data were measured on a MAT 8500 Finnigan 70 eV mass spectrometer (Germany).

General Procedure for Synthesis of 2-[N-(2,4,6-Tribromophenylamido)]propanoic Acid

A solution of succinic anhydride (5 mmol) in HOAc (300 ml) was added to a solution of 2,4,6-tribromoaniline (5 mmol) in HOAc (150 ml) and the mixture was stirred overnight at room temperature. The pale yellow precipitates were filtered,

washed with cold distilled H_2O (200 ml) and air dried.

General Procedure for Synthesis of Organotin(IV) Complexes

2-[N-(2,4,6-Tribromophenylamido)]propanoic acid (1 mmol) was suspended in dry toluene (100 ml) and treated with Et_3N (0.29 ml, 1 mmol). The mixture was refluxed for 2-3 h. To this stirring solution, diorganotin dichloride (0.5 mmol) or triorganotin chloride (1 mmol) was added as solid and the reaction mixture was refluxed for 8-10 h. The reaction mixture contained Et_3NHCl was filtered off. The solvent was removed through rotary apparatus. The mass left behind was recrystallized from CHCl_3 and *n*-hexane (1:1). In case of Oct_2SnL_2 , 2-[N-(2,4,6-tribromophenylamido)]propanoic acid (1 mmol) was refluxed with Oct_2SnO (0.5 mmol) in toluene (100 ml) using Dean and Stark apparatus for removal of H_2O formed during the reaction. The remaining procedure was followed as for others.

RESULTS AND DISCUSSION

All the complexes are soluble in most of organic solvents. The synthetic pathway to the target compounds (1)-(6) are

Table 1. Physical Data for R_2SnL_2 and R_3SnL

Compound No.	Molecular formula	Mol. Wt.	M.P ($^\circ\text{C}$)	yield (%)	Elemental analysis		
					Calcd. (found)		
					%C	%H	%N
HL	$\text{C}_{10}\text{H}_8\text{NO}_3\text{Br}_3$	430	87	80	27.90 (27.81)	1.86 (1.80)	3.25 (3.10)
(1)	$\text{C}_{22}\text{H}_{20}\text{N}_2\text{O}_6\text{SnBr}_6$	1007	67	77	26.21 (26.15)	1.98 (1.91)	2.78 (2.72)
(2)	$\text{C}_{28}\text{H}_{32}\text{N}_2\text{O}_6\text{SnBr}_6$	1091	75	70	30.79 (30.68)	2.93 (2.88)	2.56 (2.47)
(3)	$\text{C}_{36}\text{H}_{48}\text{N}_2\text{O}_6\text{SnBr}_6$	1203	123	70	35.91 (35.82)	3.99 (3.90)	2.32 (2.25)
(4)	$\text{C}_{13}\text{H}_{16}\text{NO}_3\text{SnBr}_3$	593	101	78	26.30 (26.13)	2.69 (2.57)	2.36 (2.22)
(5)	$\text{C}_{22}\text{H}_{34}\text{NO}_3\text{SnBr}_3$	719	111	68	36.71 (36.62)	4.72 (4.68)	1.94 (1.89)
(6)	$\text{C}_{28}\text{H}_{22}\text{NO}_3\text{SnBr}_3$	779	95	70	43.13 (43.06)	2.82 (2.79)	1.79 (1.71)

Spectroscopic Studies of Biologically Active Organotin(IV)



R = Me (1), *n*-Bu (2)



R = Me (4), *n*-Bu (5), Ph (6)



R = *n*-Oct (3)

Scheme 1

outlined in Scheme 1. The physical data for the complexes are reported in Table 1.

Infrared Spectroscopy

The infrared spectra of di- and triorganotin complexes were recorded in the range of 4000-400 cm^{-1} as KBr disc. The IR absorption bands for structural assignment are given in Table 2. The complexation of tin with ligand is confirmed by the presence of Sn-O band in range of 427-408 cm^{-1} which was absent in the ligand. Bands in the range of 536-528 cm^{-1} indicate the presence of Sn-C in these compounds. The bands for -NH and -OH are observed almost in the same range but these are distinguished by broad band for -OH and sharp band for -NH group in the ligand [10]. The difference between $\nu_{asym}COO - \nu_{sym}COO = \Delta\nu$ is very important in prediction of nature of ligand. In all complexes the difference $\Delta\nu$ is less than 200 cm^{-1} which indicates the bidentate nature of the ligand in all complexes [11,12].

Referring to the literature [13], the geometry of tin atom in the diorganotin(IV) carboxylates is based on skew-trapezoidal

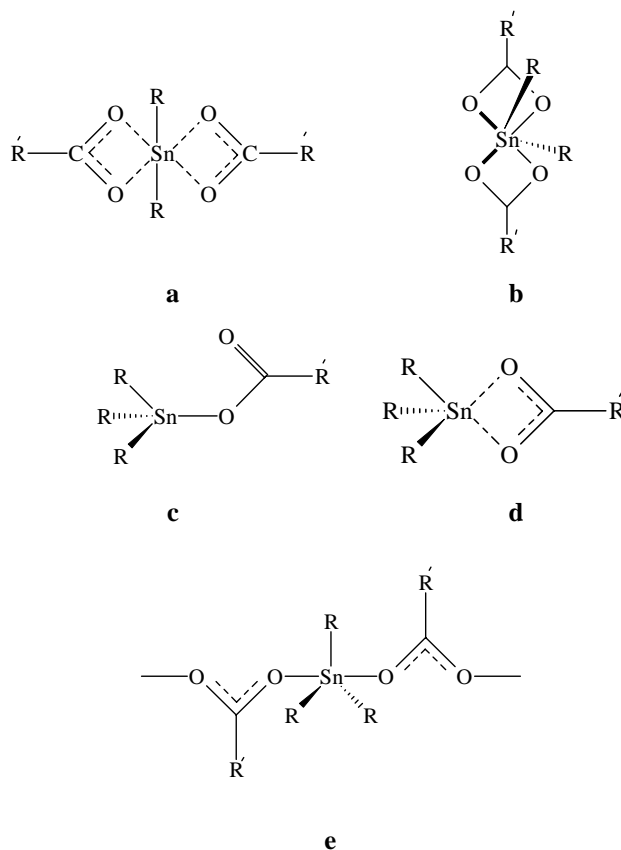


Fig. 2. Proposed structures (a), (b) for diorganotin(IV) derivatives and (c), (d), (e) for triorganotin(IV) carboxylates.

bipyramidal geometry shown in Figs. 2a and 2b. The triorganotin(IV) carboxylates are known to adopt a variety of motifs in the solid state, Figs. 2c-2e [13,14]. However, it is likely that the triorganotin(IV) species are linear polymers

Table 2. IR Spectral Data for R_2SnL_2 and R_3SnL (cm^{-1})

Compound	ν_{NH}	$\nu_{C=O}$	ν_{COO} (asym)	ν_{COO} (sym)	$\Delta\nu$	ν_{Sn-C}	ν_{Sn-O}
HL	3329	1765	1590	1346	244	-	-
(1)	3332	1760	1590	1432	158	533	411
(2)	3326	1764	1570	1410	160	528	422
(3)	3331	1769	1580	1416	164	531	408
(4)	3325	1766	1596	1411	185	530	416
(5)	3321	1770	1570	1431	139	536	427
(6)	3323	1772	1580	1432	148	-	418

(Fig. 2e) as commonly found for triorganotin(IV) carboxylates with bidentate ligands leading to *trans*-R₃SnO₂ geometry for tin [13]. In conclusion, the infrared data ($\Delta\nu$) suggest a bidentate coordination of COO group to tin atom for carboxylate ligands in the solid state.

Mass Spectrometry

Molecular ion peaks were not observed. It appears that bond dissociation energies are relatively low so that they suffer considerable fragmentation. In diorganotin(IV) derivatives, the primary fragmentation is due to the loss of ligand, R''COO. While the secondary fragmentation occurs through elimination of hydrogen (H) or CO₂ or simply involves loss of ligand (R''COO) where R'' = C₉H₇Br₃NO. Tertiary fragmentation occurs *via* cleavage of ligand or the ligand moiety R' and gives the fragments [RSnR'']⁺ and [R₂Sn]⁺, respectively, and is followed by elimination of R or R'' and ending at [Sn]⁺ (*m/z* = 120).

In triorganotin(IV) carboxylates, the primary fragmentation is due to the loss of R''COO group, followed by successive cleavage of R groups and ends at [Sn]⁺. A second fragmentation pathway is characterized by the loss of R group in primary fragmentation followed by liberation of CO₂. The secondary and tertiary fragmentations involve the loss of R or R' and end at [Sn]⁺ [15,16]. This route is the most probable than the first one. The most common fragments together with their *m/z* ratios and relative abundances is given in Table 3.

NMR Spectroscopy

The ¹H NMR chemical shifts and coupling constants for reported compounds (1)-(6) are given in Table 4. The -NH resonance appears at 4.59 ppm for all the complexes. The aromatic protons give signals in the range of 7.28-7.69 ppm. The different R groups attached to tin atom give signals at expected range thus confirming the complexation. The values of coupling constant 50 and 75 Hz for di- and triorganotin derivative, respectively, provides information regarding coordination number [17] and organotin(IV) structure.

The ¹³C NMR data also reveals expected signals with specified range. The presence of -CO and -COO was confirmed by the signals in the range of 164.2-159.9 and 177-179 ppm, respectively. The R groups attached to tin are in expected range which are reported in Table 5. The value of

Sn-C coupling ⁿJ[¹¹⁹Sn-¹³C] describes a tetrahedral geometry around the tin atom. The magnitudes for ⁿJ[¹¹⁹Sn, ¹³C] coupling are also observed and are given in Table 6. The coupling constants, ⁿJ[¹¹⁹Sn, ¹³C] are important parameters for the determination of C-Sn-C bond angles and structure characterization of organotin(IV) compounds. Coupling parameter (*J*) can easily be measured in solution, while θ can be calculated by using the Lockhart's equation (4) [18].

$$\theta = 0.0161 [{}^2J]^2 - 1.32 [{}^2J] + 133.4 \quad (4)$$

This equation is used for non-coordinating solvents. Similarly on substituting value of θ in the Lockhart's equation (5) [18], ¹J[¹¹⁹Sn, ¹³C] can be calculated.

$${}^1J[{}^{119}\text{Sn}, {}^{13}\text{C}] = 11.4 \theta - 875 \quad (5)$$

In order to gain the further information about the possible coordination geometries in solution, a close examination of the ¹J[¹¹⁹Sn-¹³C] and ²J[¹¹⁹Sn-¹H] coupling constants were undertaken, as structural details, such as the determination of C-Sn-C bond angles, can be enumerated by use of the literature methods [18,19]. For triorganotin compounds the magnitudes of ¹J[¹¹⁹Sn, ¹³C] coupling suggest the typical tetrahedral geometry around the tin atom in solution [20], while diorganotin dicarboxylates in non-coordinating solvents may acquire penta-coordinated geometry around the tin atom [20].

The chemical shifts of ¹¹⁹Sn are found in the range of -108.7 to 154.3 ppm for all complexes. For diorganotin dicarboxylates, it suggests a coordination number of tin greater than five due to increased polarization of phenyl groups. The ¹¹⁹Sn chemical shifts move to higher field [21] as the electron releasing power of the alkyl group increases the tin atom becomes progressively more shielded and ¹¹⁹Sn chemical shifts value moves to higher field. The ¹¹⁹Sn chemical shifts for compound (4) and (5) are 101.2 and 154.3, respectively [22-25].

The ¹¹⁹Sn chemical shifts for triorganotin carboxylates also support the geometry on the basis of ¹H and ¹³C NMR data as reported in Table 7. The ¹¹⁹Sn chemical shift values obtained for the triorganotin(IV) derivatives lie in the range expected for a tetrahedral geometry whereas the diorganotin(IV)

Spectroscopic Studies of Biologically Active Organotin(IV)

Table 3. Mass Spectral Data for R₂SnL₂ and R₃SnL

Fragment ion	(1)	(2)	(3)	(4)	(5)	(6)
[R ₃ Sn] ⁺	-	-	-	165(16)	291(11)	351(16)
[R ₂ Sn] ⁺	150(12)	234(15)	346(8)	150(9)	234(4)	274(10)
[BrC ₇ H ₅ OSn] ⁺	304(10)	304(3)	304(4)	304(6)	304(10)	304(5)
[BrC ₆ H ₅] ⁺	155(6)	155(12)	155(7)	155(4)	155(9)	155(100)
[Br ₃ C ₆ H ₅ N] ⁺	328(100)	328(100)	328(100)	328(100)	328(100)	328(92)
[C ₇ H ₇] ⁺	-	-	-	-	-	-
[C ₆ H ₅] ⁺	77(18)	77 (16)	77 (57)	77 (18)	77 (1)	77 (18)
[Sn] ⁺	120(5)	120(6)	120(3)	120(2)	120(7)	120(6)

Table 4. ¹H NMR Data^a (ppm) for R₂SnL₂ and R₃SnL

Compound	Br ₃ -C ₆ H ₂	-NH	-CH ₂ -CH ₂ -	R
HL	7.52s	4.59s	8.02-8.08d (9.1) 8.42-8.51d (9.1)	-
(1)	7.28s	4.59s	8.12-8.16d (9.1) 8.39-8.43d (9.1)	1.28s [74]
(2)	7.53s	4.59s	8.11-8.18d (9.1) 8.36-8.40d (9.1)	0.96t, 1.25-1.33m
(3)	7.53s	4.59s	8.09-8.14d (9.1) 8.34-8.49d (9.1)	0.89t [50], 1.68-1.75m
(4)	7.55s	4.59s	8.05-8.20d (9.1) 8.31-8.45d (9.1)	0.73s [58]
(5)	7.49s	4.59s	8.08-8.19d (9.1) 8.34-8.46d (9.1)	0.94t [75], 1.15-1.69m
(6)	7.69s	4.59s	8.02-8.18d (9.1) 8.37-8.48d (9.1)	7.48-7.84m

^aChemical shifts (δ) in ppm. ²J(¹¹⁹Sn, ¹H) and ³J(¹H, ¹H) in Hz are listed in square brackets and parenthesis, respectively. Multiplicity is given as: s = singlet, d = doublet, t = triplet, m = multiplet.

compounds indicate higher coordination, number [21].

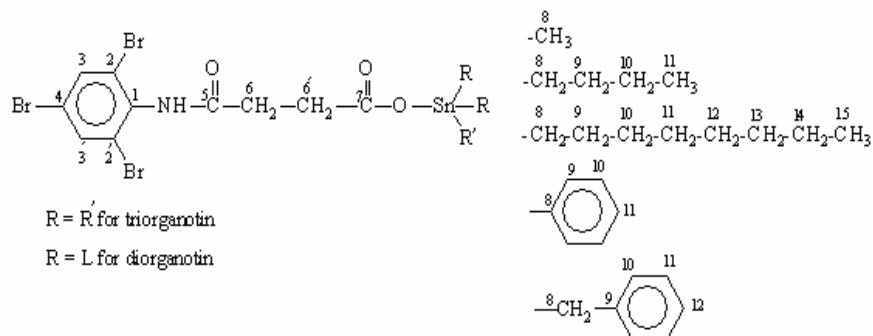
Biological Activity

Bioactive compounds are often toxic to Shrimp larvae and *in vivo* lethality to these species can be used as a rapid and simple preliminary monitor for bioactive compounds during the isolation of natural products. Brine Shrimp's method [26] has been used for the determination of toxicity of the organotin carboxylates. The results are reported in Table 8. As it is seen, compound (4) shows the highest toxicity with LD₅₀ value 10.99 μg ml⁻¹, while the other compounds also show

some toxicity. The toxicity of organotin compounds depends upon the nature of organic group as earlier reports manifested [27].

All synthesized compounds were also screened for their antibacterial activity by agar well diffusion method [28]. The results have been reported in Table 9. All compounds show significant antibacterial activity against *Escherichia coli*, *Bacillus subtilis*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Salmonella typhi* and *Shigella flexneri*.

Antifungal activity data is given in Table 10 and the tube diffusion method [28] is used. Earlier reports show that higher

Table 5. ^{13}C NMR Data (ppm) for R_2SnL_2 and R_3SnL 

^{13}C No.	HL	(1)	(2)	(3)	(4)	(5)	(6)
1	146.2	146.2	147.1	147.5	147.5	147.6	147.4
2,2'	133.7	133.7	133.4	133.7	133.7	133.7	133.9
3,3'	108.8	108.8	108.9	108.7	108.7	108.7	108.8
4	141.3	141.3	141.5	141.2	141.2	141.3	142.9
5	162.5	162.5	159.9	164.2	162.1	160.2	164.0
6,6'	128.3	128.3	125.0	129.1	128.7	128.7	128.8
7	172.0	178.0	177.0	178.0	179.0	177.0	179.0
8	-	6.8 (536)	25.5 (517,544)	26.6 (476)	-1.41 (382,394)	17.4 (345,365)	138.1
9	-	-	26.6 (21)	25.4 (31)	-	28.6 (21)	136.5
10	-	-	26.3 (69)	34.2 (92)	-	27.8 (65)	130.4
11	-	-	14.1	29.9	-	13.6	125.2
12	-	-	-	29.4	-	-	-
13	-	-	-	32.6	-	-	-
14	-	-	-	24.6	-	-	-
15	-	-	-	14.1	-	-	-

Chemical shifts (δ) in ppm. $^nJ[^{117/119}\text{Sn}, ^{13}\text{C}]$; $^nJ[^{119}\text{Sn}, ^{13}\text{C}]$ in Hz are listed in parenthesis.

Table 6. (C-Sn-C) angles ($^\circ$) Based on NMR Parameters of Organotin(IV) Derivatives for R_2SnL_2 R_3SnL

Compound No.	Compound	$^1J[^{119}\text{Sn}, ^{13}\text{C}]$ (Hz)	$^2J[^{119}\text{Sn}, ^1\text{H}]$ (Hz)	Angle ($^\circ$)	
				1J	2J
(1)	Me_2SnL_2	536	74	123.8	124.0
(3)	$n\text{-Oct}_2\text{SnL}_2$	476	-	122.4	-
(4)	Me_3SnL	394	58	111.4	111.3
(5)	$n\text{-Bu}_3\text{SnL}$	365	-	111.3	-

Table 7. ^{119}Sn NMR Data (ppm) for R_2SnL_2 and R_3SnL

Compound	(1)	(2)	(3)	(4)	(5)	(6)
^{119}Sn δ (ppm)	-113.0	-142.2	-144.2	101.2	154.3	-108.7

Spectroscopic Studies of Biologically Active Organotin(IV)

Table 8. Cytotoxicity Data for R₂SnL₂ and R₃SnL

Compound No.	LD ₅₀
(1)	16.19
(2)	7.88
(3)	8.88
(4)	10.99
(5)	8.99
(6)	9.9

antifungal activity is associated with tributyltin and triphenyltin compounds [29]. Tributyltin carboxylate shows highest activity against *Trichophyton longiusus*, *Aspergillus flavis*, *Microsporium canis*, *Fusarium solani* and *Candia glaberata*, while dibutyltin and trimethyltin carboxylates do not show any activity.

The insecticidal activity data of the compounds are given in Table 11 and the data were collected by contact toxicity

Table 9. Antibacterial Activity for R₂SnL₂ and R₃SnL

Name of bacteria	Zone of inhibition (mm)					
	(1)	(2)	(3)	(4)	(5)	(6)
<i>Escherichia coli</i>	12	-	14	15	-	14
<i>Bacillus subtilis</i>	14	-	14	10	-	15
<i>Shigella flexenari</i>	16	-	18	8	-	15
<i>Staphylococcus aureus</i>	16	-	8	8	5	15
<i>Pseudomonas aeruginosa</i>	16	-	12	12	-	10
<i>Salmonella typhi</i>	10	-	14	18	-	16

Standard drug = Ampicilline (H₂O)₃, Cephalexin Na. Concentration of standard drug = 3 mg ml⁻¹.

Table 10. Antifungal Activity for R₂SnL₂ and R₃SnL

Fungi	Compound No.							Standard drug
	HL	(1)	(2)	(3)	(4)	(5)	(6)	
<i>Trichophyton longiusus</i>	89.4	88.8	-	100	-	100	55	Miconazole Ketoconazole
<i>Candida albicans</i>	-	-	-	100	-	100	-	Miconazole Ketoconazole
<i>Aspergillus flavis</i>	-	-	-	100	-	100	-	Amphotericin-B Flucytosine
<i>Microsporium canis</i>	-	-	-	100	-	100	-	Miconazole Ketoconazole
<i>Fusarium solani</i>	-	-	-	100	-	100	-	Miconazole
<i>Candida glaberata</i>	-	-	-	100	-	100	-	Miconazole

Table 11. Insecticidal Data for R₂SnL₂ and R₃SnL

Insects	Compounds					
	(1)	(2)	(3)	(4)	(5)	(6)
<i>Tribolium continuum</i>	-	-	-	-	-	25
<i>Sitophilus oryzae</i>	-	-	25	-	-	-
<i>Rhyzopertha dominica</i>	25	-	25	-	25	25
<i>Callosbruchus analis</i>	-	-	25	-	-	-

Standard drug = Permethrin. Concentration of standard drug = 235.55 µg cm⁻².

Table 12. Antileishmanial Activity for R₂SnL₂ and R₃SnL

Compound No.	IC ₅₀
(1)	61.00
(2)	65.15
(3)	62.50
(4)	62.00
(5)	61.00
(6)	58.00

Standard drug = Amphotericin. Concentration of standard drug = 0.19 µg ml⁻¹.

method [28]. It shows that compound (1) is found to be inactive against the tested insects while the compounds (3), (5) and (6) show the activity.

Leishmaniasis is caused by a protozoa species of the genus leishmania. The organotin derivatives were tested for antileishmanial activity and was found to be significantly more active in reducing the parasite load at much lower concentration (Table 12). The results suggest that the compounds could be exploited as an antileishmanial drug and have potential to be used as a new agent for leishmaniasis.

Thus the presence of an organic group directly attached to tin is an important factor which is responsible for enhanced activity of organotin(IV) complexes. This indicates that the alkyl group directly appended to the central tin atom is important contributor to the activity.

CONCLUSIONS

It is observed that the multinuclear NMR data for all complexes found were in agreement with reported values. IR spectroscopy proved the bidentate nature of the carboxylic groups and suggests the penta and hexa coordinated geometry in solid state for tri- and diorganotin derivatives, respectively. Mass spectral data are also in agreement with the proposed molecular formulae of all the synthesized compounds.

A particular feature of these complexes is that the properties of central metal atom may be tuned by introducing different substituents particularly in regard to their biological activity.

ACKNOWLEDGEMENTS

SA is thankful to Pakistan Science Foundation for financial support under the Project No. PSF/R & D/C-QU/Chem (270).

REFERENCES

- [1] A.G. Davis, P.G. Smith, in: G. Wilkinson, F.G.A. Stone, E.W. Abel (Eds.), *Comprehensive Organometallic Chemistry*, Pergamon, Oxford, UK, 1982.
- [2] M. Pereyer, J.P. Quintard, A. Rahim, *Tin in Organic Synthesis*, Butterworth, London, 1987.
- [3] D. Martin, U. Russo, D. Stivanello, G. Tagliavini, *Organometallics* 15 (1996) 1645.
- [4] G.S. Drummond, A. Kappass, *Proc. Natl. Acad. Sci.* 78 (1981) 6466.
- [5] K.C. Molloy, T.G. Purcell, K. Quil, I.W. Nowell, *J. Organomet. Chem.* 267 (1984) 237.
- [6] K.C. Molloy, T.G. Purcell, E. Mahn, H. Schumann, J.J. Zuckerman, *Organometallics* 5 (1986) 85.
- [7] A. Ruzicka, L. Dostal, R. Jambor, V. Buchta, J. Brus, I. Cisarova, M. Holcapek, J. Holecek, *Appl. Organomet. Chem.* 16 (2002) 315.
- [8] K. Shahid, S. Ali, S. Shahzadi, A. Badshah, K.M. Khan, G.M. Maharvi, *Synth. React. Inorg. Met.-Org. Chem.* 33 (2003) 1221.
- [9] S. Shahzadi, K. Shahid, S. Ali, M. Mazhar, K.M. Khan, *J. Iran. Chem. Soc.* 2 (2005) 277.
- [10] M. Gielen, A. El-Khaloufi, F. Kayrev, M. Biesemans, R. Willem, *Appl. Organomet. Chem.* 7 (1993) 201.
- [11] Q.L. Xie, Z.Q. Yang, Z.X. Zhang, D.K. Zhang, *Appl. Organomet. Chem.* 6 (1992) 193.
- [12] Q. Xie, Z. Yang, L. Jiang, *Main Group Met. Chem.* 19 (1996) 509.
- [13] Sadiq-ur-Rehman, S. Ali, A. Badshah, A. Malik, E. Ahmed, G.-X. Jin, E.R.T. Tiekink, *Appl. Organomet. Chem.* 18 (2004) 401.
- [14] Sadiq-ur-Rehman, K. Shahid, S. Ali, M. Mazhar, A. Badshah, G. Eng, X. Song, J. Ryzkowski, *Heteroatom Chem.* 16 (2005) 175.
- [15] A. Badshah, M. Danish, S. Ali, M. Mazhar, S. Mahmood, M.I. Chaudhry, *Synth. React. Inorg. Met.*

Spectroscopic Studies of Biologically Active Organotin(IV)

- Org. Chem. 24 (1994) 1155.
- [16] T.N. Metcchell, J. Organomet. Chem. 59 (1973) 189.
- [17] B. Jousseume, J.G. Duboudin, M. Petraud, J. Organomet. Chem. 238 (1982) 171.
- [18] T.P. Lockhart, W.F. Manders. Inorg. Chem. 25 (1986) 892.
- [19] T.P. Lockhart, W.F. Manders, E.M. Holts, J. Am. Chem. Soc. 108 (1986) 6611.
- [20] S. Ali, F. Ahmad, M. Mazhar, A. Munir, M.T. Masood, Synth. React. Inorg. Met. -Org. Chem. 32 (2002) 357.
- [21] B. Wrackmeyer, G. Kehr, J. Süß Chem. Ber. 126 (1993) 2221.
- [22] K. Shahid, S. Ali, S. Shahzadi, Z. Akhtar, Turk. J. Chem. 27 (2003) 209.
- [23] K. Shahid, S. Shahzadi, S. Ali, M. Mazhar, Bull. Korean Chem. Soc. 27 (2006) 44.
- [24] S. Ahmad, S. Ali, S. Shahzadi, F. Ahmed, K.M. Khan, Turk. J. Chem. 29 (2005) 299.
- [25] Sadiq-ur-Rehman, S. Ali, S. Shahzadi, A. Malik, E. Ahmad, Turk. J. Chem. 31 (2007) 371.
- [26] J.M. Barnes, H.B. Stoner, Brit. J. Ind. Med. 15 (1958) 15.
- [27] H. Blank, G. Rewbell, Arch. Derm. 92 (1965) 319.
- [28] A. Rahman, M.I. Choudhry, W.J. Thomsen, Bioassay Techniques for Drug Development, Harward Academic Press, Amsterdam, 2001.
- [29] S.S. Shaukat, N.A. Khan, F. Ahmed, Pak. J. Bot. 12 (1980) 97.